

THE STRUCTURE OF A URINARY METABOLITE OF PROSTAGLANDIN  $E_2$   
IN THE GUINEA PIG

Mats Hamberg and Bengt Samuelsson

Department of Medical Chemistry, Royal Veterinary College,  
Stockholm, Sweden

Received November 12, 1968

The excretory products of a number of prostaglandins in the rat have been characterized. Thus, one of the urinary metabolites of  $5,6\text{-}^3\text{H}_2\text{-PGF}_{1\alpha}$ <sup>1</sup> was identified as dinor-PGF<sub>1 $\alpha$</sub>  (Granström *et al.* 1965), whereas a urinary metabolite formed from  $9\beta\text{-}^3\text{H-PGF}_{2\alpha}$  was tetranor-PGF<sub>1 $\alpha$</sub>  (Gréen and Samuelsson 1968). The structures of the urinary metabolites of PGE<sub>1</sub> have not been determined. However, the labeled material excreted in the urine after administration of  $5,6\text{-}^3\text{H}_2\text{-PGE}_1$  consisted exclusively of compounds more polar than PGE<sub>1</sub> (Samuelsson 1964) and experiments with  $3\text{-}^{14}\text{C-PGE}_1$  indicated that the formation of these metabolites involved degradation of the carboxyl side chain of PGE<sub>1</sub> by four carbon atoms (Samuelsson 1965). In accordance with these findings the  $^{14}\text{C}$  label of  $1\text{-}^{14}\text{C-PGE}_1$  administered to rats was mainly recovered in the expired carbon dioxide (Miller and Krake 1968).

---

1 The following trivial names and abbreviations are used: Prostaglandin  $E_1$  (PGE<sub>1</sub>), 11 $\alpha$ ,15-dihydroxy-9-ketoprost-13-enoic acid; prostaglandin  $E_2$  (PGE<sub>2</sub>), 11 $\alpha$ ,15-dihydroxy-9-ketoprost-5,13-dienoic acid; prostaglandin  $B_2$  (PGB<sub>2</sub>), 15-hydroxy-9-ketoprost-5,8(12),13-trienoic acid; prostaglandin  $F_{1\alpha}$  (PGF<sub>1 $\alpha$</sub> ), 9 $\alpha$ ,11 $\alpha$ ,15-trihydroxyprost-13-enoic acid; prostaglandin  $F_{1\beta}$  (PGF<sub>1 $\beta$</sub> ), 9 $\beta$ ,11 $\alpha$ ,15-trihydroxyprost-13-enoic acid; and prostaglandin  $F_{2\alpha}$  (PGF<sub>2 $\alpha$</sub> ), 9 $\alpha$ ,11 $\alpha$ ,15-trihydroxyprost-5,13-dienoic acid.

The present communication describes experiments which have led to identification of the major urinary metabolite of  $\text{PGE}_2$  in the guinea pig as  $5\beta,7\alpha$ -dihydroxy-11-ketotetranor-prostanoic acid.

#### METHODS AND RESULTS

$17,18\text{-}^3\text{H}_2\text{-PGE}_2$  (1.5-1000  $\mu\text{g}$ ), prepared as previously described (Änggård *et al.* 1965) and with a specific activity of 420 or 0.42  $\mu\text{C}/\mu\text{mole}$ , was injected subcutaneously to male guinea pigs. Of the injected radioactivity 35-40% was recovered in the urine within 12 hours. Of the labeled material present in the urine 85-90% could be extracted with diethyl ether after acidification. The extracted material was fractionated by reversed phase partition chromatography (column 9 g of hydrophobic Hyflo Super-Cel, solvent system C-36, c.f. Hamberg 1968). The material forming the main peak of radioactivity (Metabolite I, 96-120 ml effluent) was subjected to silicic acid chromatography (eluted with ethyl acetate - benzene, 80:20). The methyl ester, prepared by treatment with diazomethane solution, was again subjected to silicic acid chromatography (eluted with ethyl acetate - benzene, 60:40). This procedure yielded material which behaved homogenously upon thin layer chromatography (solvent system M II, c.f. Gréen and Samuelsson 1964,  $R_f = 0.75$ ; reference  $\text{PGE}_1$  methyl ester,  $R_f = 0.83$ ) and gas-liquid chromatography (see below).

On treatment of the methyl ester of Metabolite I with sodium hydroxide no absorption band at 237  $\text{m}\mu$  or 278  $\text{m}\mu$  developed. Under the conditions used,  $\text{PGE}_2$  was transformed into  $\text{PGB}_2$  ( $\lambda_{\text{max}}^{\text{EtOH}} = 278 \text{ m}\mu$ ) and  $11\alpha,15$ -dihydroxy-9-ketoprostanoic acid was transformed into 15-hydroxy-9-ketoprost  $\sim 8(12)$ -enoic acid ( $\lambda_{\text{max}}^{\text{EtOH}} = 237 \text{ m}\mu$ ) (c.f. Bergström *et al.* 1963).

Gas chromatographic-mass spectrometric analysis of four derivatives of the methyl ester of Metabolite I (c.f. the Table and Fig. 3) indicated the following structural features: carbon chain of 16 carbon atoms, two hydroxyl groups and one keto group. The two hydroxyl groups were

Table. Retention times on gas-liquid chromatography<sup>x</sup>.

Derivative	1% SE 30			1% EGSS-X		
	(1) <sup>xx</sup>	(2) <sup>xx</sup>	(3) <sup>xx</sup>	(1) <sup>xx</sup>	(2) <sup>xx</sup>	(3) <sup>xx</sup>
TMSi	21.0	20.7	20.7	23.3	22.8	22.8
TMSi-MO	21.1	20.8	20.8	22.7	22.3	22.3
Acetate	21.9	21.8	21.8			
Acetate-MO	22.0	21.9	21.9			

TMSi, trimethylsilyl ether; MO, O-methyl oxime.

x) C-values, see Hamberg 1968

xx) (1) methyl 5 $\alpha$ ,7 $\alpha$ -dihydroxy-11-ketotetranor-prostanoate; (2) methyl 5 $\beta$ ,7 $\alpha$ -dihydroxy-11-ketotetranor-prostanoate; (3) methyl ester of Metabolite I.

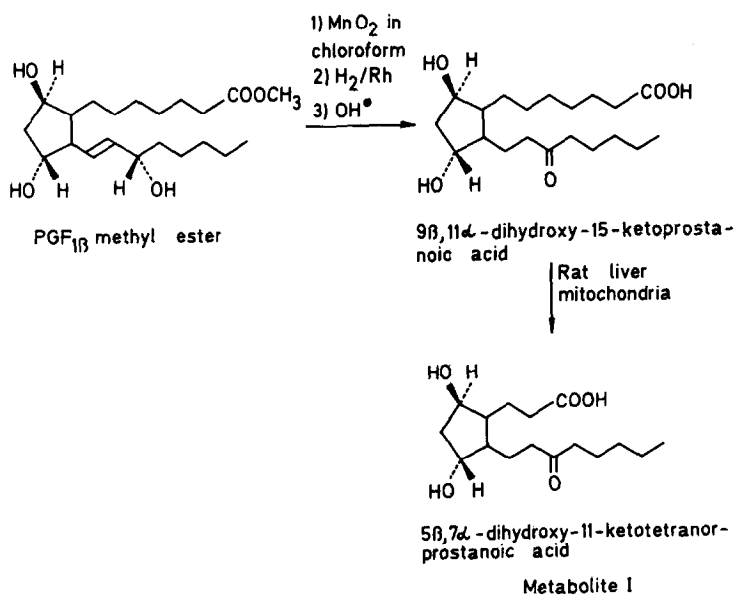


Fig. 1. Reactions used to prepare 5 $\beta$ ,7 $\alpha$ -dihydroxy-11-ketotetranor-prostanoic acid.

most likely located in the five-membered ring and the keto group in the side chain, since ultraviolet spectrometry indicated that the  $\beta$ -ketol system of  $\text{PGE}_2$  was not retained in Metabolite I. The retention times (expressed as C-values, c.f. Hamberg 1968) of the trimethylsilyl ether, trimethylsilyl ether- O-methyl oxime, acetate, and acetate- O-methyl oxime derivatives prepared of the methyl ester of Metabolite I were identical with those of the corresponding derivatives of methyl  $5\beta,7\alpha$ -dihydroxy-11-ketotetranor-prostanoate using 1% SE 30 and 1% EGSS-X (see the Table).

For the preparation of  $5\alpha,7\alpha$ -dihydroxy-11-ketotetranor-prostanoic acid and  $5\beta,7\alpha$ -dihydroxy-11-ketotetranor-prostanoic acid,  $\text{PGF}_{1\alpha}$  and  $\text{PGF}_{1\beta}$  were oxidized and hydrogenated (c.f. Änggård and Samuelsson 1964) to give  $9\alpha,11\alpha$ -dihydroxy-15-ketoprostanoic acid and  $9\beta,11\alpha$ -dihydroxy-15-ketoprostanoic acid. These compounds were then converted into the corresponding  $\text{C}_{16}$  homologues using the  $\beta$ -oxidation system of rat liver (Hamberg 1968) (Fig. 1).

The mass spectra of the four derivatives of the methyl ester of Metabolite I were identical with those of the corresponding derivatives of methyl  $5\beta,7\alpha$ -dihydroxy-11-ketotetranor-prostanoate. The mass spectra of the trimethylsilyl ether derivatives are shown in Fig. 2 and Fig. 3. Ions of high intensity are present at  $m/e$  443 (M-15), 427 (M-31), 368 (M-90), 353 (M-(15+90)), 281 (M-(87+90)), 278 (M-2x90), 254 (M-(114+90)), 241 (M-(127+90)), 217, 191 (M-(87+2x90)) and 179 (M-(99+2x90)) (c.f. Fig. 2).

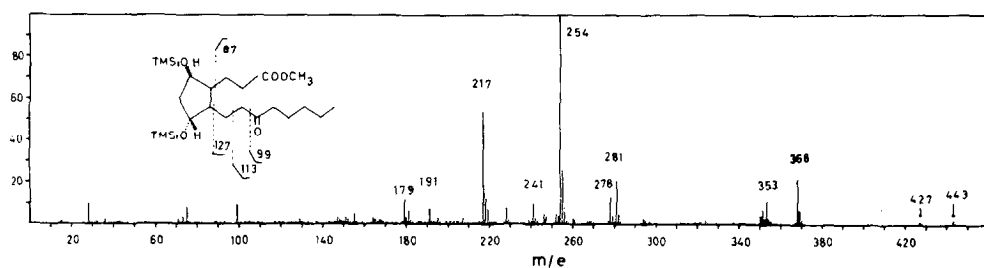


Fig. 2. Mass spectrum of trimethylsilyl ether derivative of methyl  $5\beta,7\alpha$ -dihydroxy-11-ketotetranor-prostanoate.

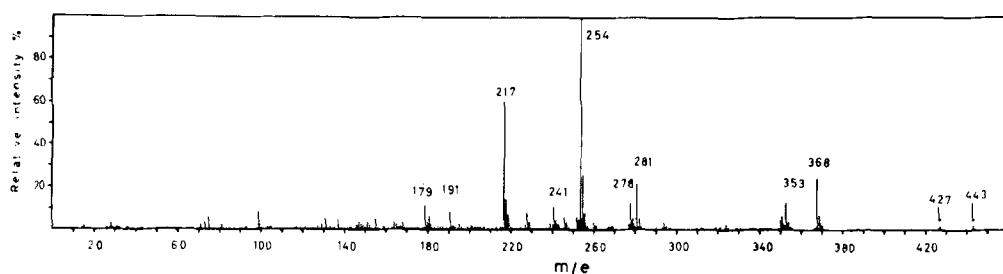


Fig. 3. Mass spectrum of trimethylsilyl ether derivative of the methyl ester of Metabolite I.

#### DISCUSSION

The formation of 5 $\beta$ ,7 $\alpha$ -dihydroxy-11-ketotetranor-prostanoic acid from PGE<sub>2</sub> involves oxidation of the secondary alcohol group in the side chain, reduction of the trans double bond,  $\beta$ -oxidation of the carboxyl side chain and reduction of the keto group in the five-membered ring. The first two reactions were originally discovered in guinea pig lung and were also demonstrated to occur in other tissues and species (Änggård and Samuelsson 1964, Änggård and Samuelsson 1967). Degradation of the carboxyl side chain of prostaglandins by  $\beta$ -oxidation has been demonstrated previously both in vivo (Granström et al. 1965, Gréen and Samuelsson 1968) and in vitro (Hamberg 1968). However, reduction of the keto group in the five-membered ring has not previously been observed. The configuration ( $\beta$ ) of the hydroxyl group formed is opposite to that found in "primary" prostaglandins. Studies on the enzyme system catalyzing this reaction as well as the sequence of the reactions involved in the transformation of PGE<sub>2</sub> into 5 $\beta$ ,7 $\alpha$ -dihydroxy-11-ketotetranor-prostanoic acid are in progress in our laboratory.

#### ACKNOWLEDGEMENTS

The authors are indebted to Mrs. G. Hamberg for expert technical assistance.

This work was supported by the Swedish Medical Research Council (proj. no. 13X-217) and by the Knut and Alice Wallenbergs Stiftelse.

## REFERENCES

- Änggård, E., and Samuelsson, B., J. Biol. Chem., 239, 4097 (1964).  
Änggård, E., Gréen, K., and Samuelsson, B., J. Biol. Chem., 240, 1932 (1965).  
Änggård, E., and Samuelsson, B., in S. Bergström and B. Samuelsson (Editors), Proceedings of the Second Nobel Symposium, Almquist and Wiksell, Stockholm, 1967, p. 97.  
Bergström, S., Ryhage, R., Samuelsson, B., and Sjövall, J., J. Biol. Chem., 238, 3555 (1963).  
Granström, E., Inger, U., and Samuelsson, B., J. Biol. Chem., 240, 457 (1965).  
Gréen, K., and Samuelsson, B., J. Lipid Res., 5, 117 (1964).  
Gréen, K., and Samuelsson, B., Presented at the 3rd Internat. Symp. on Drugs affecting Lipid Metabolism (Milano 1968).  
Hamberg, M., Eur. J. Biochem., 6, 135 (1968).  
Miller, W.L., and Krake, J.J., Fed. Proc., 27, 241 (1968).  
Samuelsson, B., J. Biol. Chem., 239, 4091 (1964).  
Samuelsson, B., Unpublished observations (1965).